



Effect of Dorsomorphin Homolog 1 (DMH1) against Diabetic Dyslipidemia in Streptozotocin-induced Diabetic Rats

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

Dyslipidemia is usually observed in both types of diabetes and, particularly, "atherogenic dyslipidemic triad" is strongly linked to a higher risk of adverse cardiovascular outcome. On the other hand, bone morphogenetic proteins (BMP) are a group of wide variety of proteins which were found overexpressed and implicated in contribution and acceleration of atherosclerotic calcification. So, the present study aimed to assess effect of DMH1, a selective BMP inhibitor, in a rat model of diabetic-induced dyslipidemia.

Methods: STZ-induced diabetes in Wistar rats was used as a model to assess the antihyperlipidemic effect of DMH1(5mg/kg) for a period of 8 weeks. Rats were divided into normal control (C=10), diabetic control (DC=10), diabetic+vehicle (DV=10) and diabetic DMH1-treated rats (DT=10). Fasting blood glucose (FBG) level was measured on weekly bases. Then, at the end of the experiment, rats were anesthetized and blood samples were collected for the determination of total cholesterol (TC), triglyceride (TG), LDL and HDL levels using the appropriate ELISA assay.

Results: FBG levels for all diabetic groups were significantly high, during the experiment period, compared to the control ($P < 0.001$). While dyslipidemia was remarkable in the diabetic non-treated

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groups, DMH1 treatment showed a significant decrease in TC ($P < 0.001$), TG ($P < 0.05$) and LDL levels ($P < 0.001$) compared to the non-treated groups (DC & DV). Concurrently, HDL levels for DT group were significantly increased compared to DC or DV groups ($P < 0.01$).

Conclusion: The present experiment showed that DMH1 possessed encouraging activity against dyslipidemia in STZ-induced diabetic rats. Our results are promoting for more interest and investigation regarding antihyperlipidemic effect of DMH1 and BMP/Smad pathway in further experimental studies.

Keywords: *Dorsomorphin homolog 1; DMH1; diabetic dyslipidemia; bone morphogenetic protein; BMP.*

1. INTRODUCTION

Cardiovascular disease (CVD) is a definite health concern which commonly involved in both types of diabetes and may represent the most common cause of death [1,2]. Despite the substantial advances in diabetic care, both absolute and relative risks of CVD among insulin-dependent diabetic (T1DM) patients remained extremely high. Although CVD risk may considerably vary based on age, sex or duration of diabetes, however, patients with early-onset (<10 years of age) type 1 diabetes are carrying about 30-fold increased risks of CVD or coronary heart disease (CHD) and approximately 7.4-times more risk of cardiovascular-related mortality [3]. Furthermore, diabetes, per se, was frequently reported to carry a comparable risk to the previous cardiovascular event in non-diabetic patients [4-6].

On the other hand, dyslipidemia is usually observed in poorly controlled T1DM and obese T2DM patients [7]. Particularly, “atherogenic dyslipidemic triad” of raised triglyceride levels, low levels of high-density lipoprotein cholesterol (HDL-C) and increase in small and dense low-density lipoprotein (LDL) particles is more tightly linked to higher CVD risk and may be a useful biomarker for risk assessment [8]. Also, the association of hypertriglyceridemia with CHD was stronger in patients with diabetes than in the general population, while lipid-lowering therapy has significantly decreased CHD risks in diabetic patients as well as non-diabetics [9]. Unfortunately, under screening and under treatment of dyslipidemia as a major modifiable risk factor for CHD in diabetic patients was numerously reported [10-12].

The most common lipid abnormality in diabetic populations is hypertriglyceridemia which occurred due to either increased hepatic secretion in T2DM or defective removal process in the absolute insulin-deficient state (T1DM) [13]. Moreover, chylomicron removal by

lipoprotein lipase (LPL) is also majorly regulated by insulin. Additionally, diabetic kidney disease (DKD) is a common microvascular complication and, substantially, involved in both dyslipidemia and CVD [14]. On the other hand, bone morphogenetic proteins (BMP) are a group of wide variety of proteins forming the subset of a larger superfamily called transforming growth factor- β (TGF- β) [15]. Interestingly, a previous study of human atherosclerotic lesions revealed detection of BMPs (BMP-2 & BMP-4) in calcified atherosclerotic plaques. The report also denoted the regulatory role of BMP in atherogenesis [16]. Additional reports have outlined the involvement of BMP signaling in vascular injury and calcification [17,18]. Moreover, *In vitro* study of human coronary artery smooth muscle cells, BMP was overexpressed and implicated in the contribution and acceleration of atherosclerotic calcification [19]. Furthermore, Derwall et al, provided further evidence of BMP activation in early atherosclerotic lesions, besides the remarkable reduction in vascular calcification achieved by inhibition of BMP signaling in atherogenic animals [20]. Additionally, hyperglycemia and diabetes were found to strongly activate BMP signaling in endothelial cells and aortic wall with a consequent increase in expression of osteogenic markers and medial calcification [21].

Dorsomorphin Homolog 1 (DMH1) is a small molecule with a promising pharmacological property that almost exclusively inhibit BMP with no off-target effects [22]. DMH1 was used as BMP pharmacological inhibitor in many experiments and displayed positive results with low toxicity and effective inhibition of BMP/Smad pathway, which make it a good choice to be tested for purpose of BMP inhibition [23-25]. Therefore, the present study is the preliminary attempt to explore the effect of selective BMP inhibitor (DMH1) in experimental animals with diabetic dyslipidemia.

2. MATERIAL AND METHODS

2.1 Chemicals and Kits

Streptozocin (STZ) (Cat. No. 1621) was purchased from Tocris Bioscience® (Bristol, UK) and supplied as crystalline solid to be dissolved in 0.1 M sodium citrate buffer (pH 4.5) just prior use. DMH1 (Dorsomorphin Homolog 1) (Cat. No. 4126) was also purchased from Tocris Bioscience® (Bristol, UK) supplied as a yellow crystalline solid. (2-Hydroxypropyl)- β -cyclodextrin (HP β CD) powder (Cat. No. OH05393) was acquired from Biosynth Carbosynth® (Compton, UK) and was freshly prepared as 12.5% solution to be used as a vehicle for DMH1. Quick DetectTM Total Cholesterol (Rat) ELISA kit (Cat No.MBS846775), rat triglyceride (TG) ELISA kit (Cat No. MBS726298), rat HDL ELISA kit (Cat No.MBS2505957), rat LDL ELISA kit (Cat No.MBS702165) were all purchased from My Bio Source (San Diego, USA).

2.2 Induction of Diabetes and Experimental Design

A total of forty male adult Wistar rats which weighed (180-220) g were used for the current study. The animals were obtained from the pharmaceutical consultation and research unit, Faculty of Pharmacy, King Abdulaziz University (KAU), Jeddah, Saudi Arabia. Feed and tap water *ad libitum* were provided. Additionally, standard animal room temperature (29-30 °C) and 12 hours of light/dark cycle were maintained during the whole study period. Following one week of acclimating to the facilities, ten random rats were marked as the control (C=10), and the remaining 30 rats were prepared for diabetes induction by intraperitoneal (i.p.) injection with Streptozotocin (STZ) 60 mg/kg. Three days later, rats with blood glucose level of ≥ 300 mg/dl were considered as diabetics [26] and were randomly divided into three different groups with ten rats each, as the following: Diabetic non-treated group (DC, n = 10) supplemented by regular food and water, Diabetic non-treated group + vehicle (DV, n = 10) injected (i.p.) with equivalent amount of vehicle (12.5% 2-hydroxypropyl- β -cyclodextrin [HP β CD]) for 8 weeks, Diabetic treated group (DT, n = 10) injected (i.p.) with 5 mg/kg DMH1 each other day (q.o.d) for 8 weeks [24,27]. HP β CD was selected based on its property of increasing drug solubility and bioavailability [28], as well as being previously and successfully used as a vehicle for DMH1 [24].

Fasting blood glucose (FBG) readings were recorded using ACCU-CHEK® Active (Roche Diagnostics GmbH, Mannheim, Germany) at the baseline and on weekly basis thereafter. At the end of the experiment, blood samples were withdrawn and centrifuged at 3000g for 10 min to obtain clear sera which were used for assessment of total cholesterol (TC), triglyceride (TG), high-density lipoproteins (HDL) and low-density lipoproteins (LDL) using appropriate enzyme-linked immunosorbent assay (rat ELISA kit). The rats were sacrificed after being anesthetized with diethyl ether.

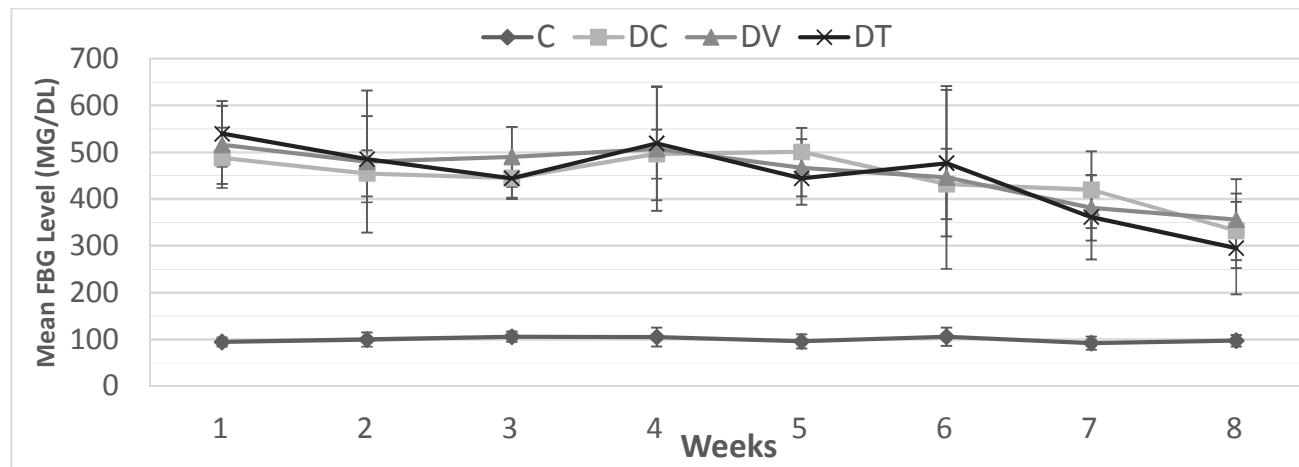
2.3 Statistics

One-way analysis of variance (ANOVA) was used to examine the gathered results using SPSS statistics software package (version 23) followed by Tukey's HSD multiple comparison post hoc test. Differences if *P*-values < 0.05 were considered statistically significant.

3. RESULTS

By the end of the experiment, a total of nine rats were dead; three rats were from the DC group, two rats were from DV group, while the other four rats were in the treated group. However, fasting blood glucose (FBG) levels were constantly and significantly high during the study period (8 weeks) for all diabetic groups (DC, DV& DT) in comparison to the control group (*P*< 0.001). Moreover, comparing FBG levels between all the three diabetic groups (DC, DV & DT) revealed no significant difference (Fig. 1).

There were remarkable differences in lipid profiles between the four different groups, as shown in Table 1. Total cholesterol (TC) & LDL levels were significantly increased for diabetic non-treated groups (DC & DV), while the treated group showed no difference versus (C) group and appeared significantly lower than DC or DV groups (*P*< 0.001) (Fig. 2, Fig. 4). Similar results were obtained for triglycerides (TG) levels for the tested groups (increased for DC & DV groups and lower levels for DT group) (*P*< 0.05) (Fig. 3). Furthermore, for the HDL levels, DT group has significantly increased levels compared to DC or DV groups (*P*< 0.01), while both DC & DV were significantly lower than the control group (*P*< 0.05). Although it was statistically insignificant (*P*> 0.05), the mean HDL for DT group was superior to that of the control group (Fig. 5).

**Fig. 1. Averages of weekly FBG levels \pm SD (mg/dl) for all groups of rats during the study period (8 weeks)**

FBG (fasting blood glucose). C = Control rats ($n=10$), DC= diabetic control group ($n=7$), DV= diabetic+vehicle group ($n=8$), DT= diabetic-DMH1 treated group [5mg/kg q.o.d] ($n=6$)

Table 1. Effect of DMH1 on lipid profile of the different groups of rats by the end of the experiment (after 8 weeks)

Parameters	Groups			
	Control	DC	DV	DT (5mg/kg)
TC (mg/dl)	123.9 \pm 11.9	257.6 \pm 50.4 **	233.6 \pm 49.4 ***	122.5 \pm 6 ***
TG (mg/dl)	74.4 \pm 7.1	126.9 \pm 40.4 *	126.6 \pm 47.5 *	76.2 \pm 3.7 #
LDL (mg/dl)	79.9 \pm 12.1	222.4 \pm 57.1 ***	201.5 \pm 53.6 ***	75.5 \pm 6.1 ***
HDL (mg/dl)	44 \pm 3	34.1 \pm 6.7 *	32.1 \pm 8 **	47.5 \pm 3.5 ##

Data were expressed as mean \pm SD. TC (Total cholesterol mg/dl). TG (Triglyceride mg/dl). LDL (low-density lipoproteins) mg/dl. HDL (high-density lipoproteins) mg/dl. C = Control rats ($n=10$), DC= Diabetic Control group ($n=7$), DV= Diabetic + Vehicle group ($n=8$), DT= Diabetic-DMH1 Treated group [5mg/kg q.o.d] ($n=6$). * significant ($P < 0.05$) against control group. ** highly significant ($P < 0.01$) against control group. *** very highly significant ($P < 0.001$) against control group. # significant ($P < 0.05$) against DC or DV group. ## highly significant ($P < 0.01$) compared to DC or DV group. ### very highly significant ($P < 0.001$) in comparison to Diabetic non-treated group (DC or DV)

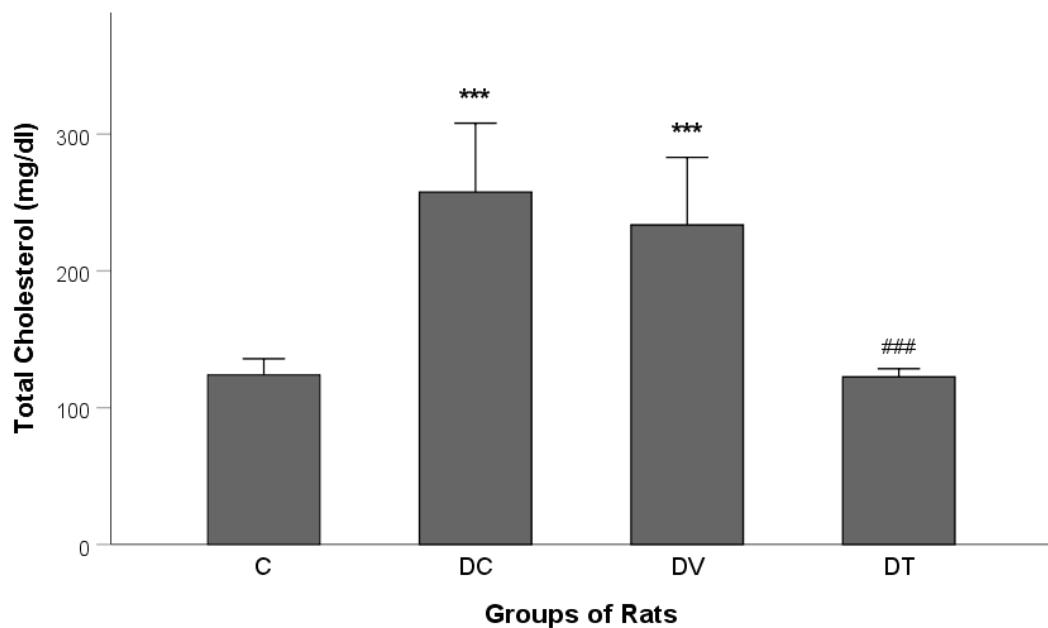


Fig. 2. Averages of total cholesterol (TC) levels for different groups of rats after 8 weeks of the study

Data were presented as mean \pm SD. TC (Total cholesterol mg/dl). C = Control rats ($n=10$), DC= Diabetic Control group ($n=7$), DV= Diabetic + Vehicle group ($n=8$), DT= Diabetic-DMH1 Treated group [5mg/kg q.o.d] ($n=6$). *** very highly significant ($P< 0.001$) against control group. ### very highly significant ($P< 0.001$) in comparison to Diabetic non-treated group (DC or DV)

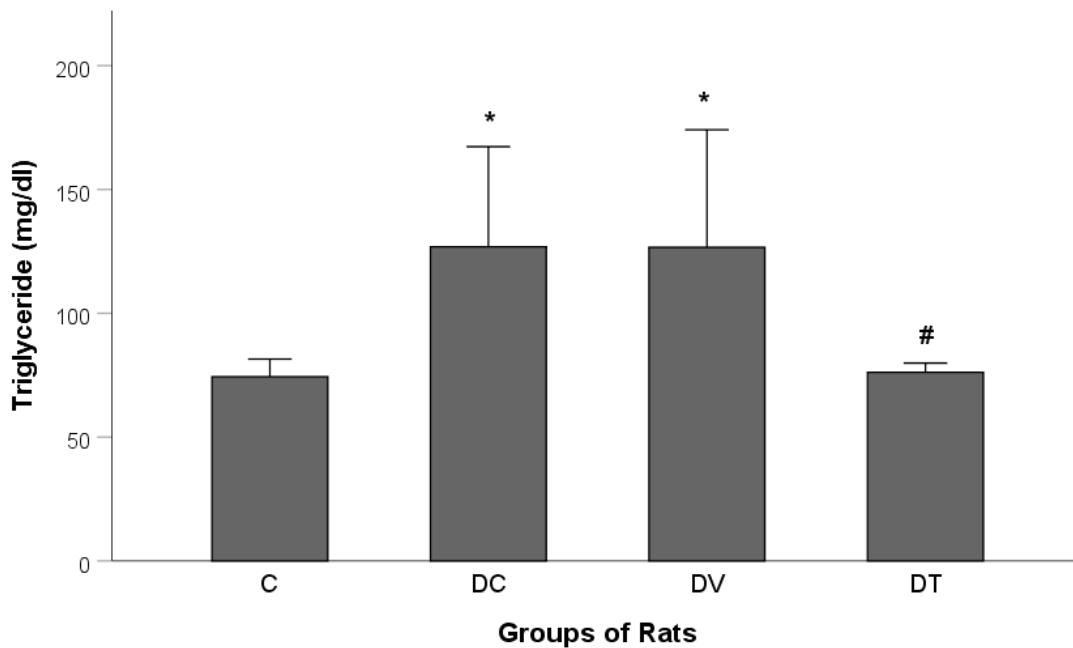


Fig. 3. Mean triglycerides (TG) levels for all tested groups of rats after 8 weeks of the study

Data were presented as mean \pm SD. TG (Triglyceride mg/dl). C = Control rats ($n=10$), DC= Diabetic Control group ($n=7$), DV= Diabetic + Vehicle group ($n=8$), DT= Diabetic-DMH1 Treated group [5mg/kg q.o.d] ($n=6$). * significant ($P< 0.05$) against control group. # significant ($P< 0.05$) against DC or DV group

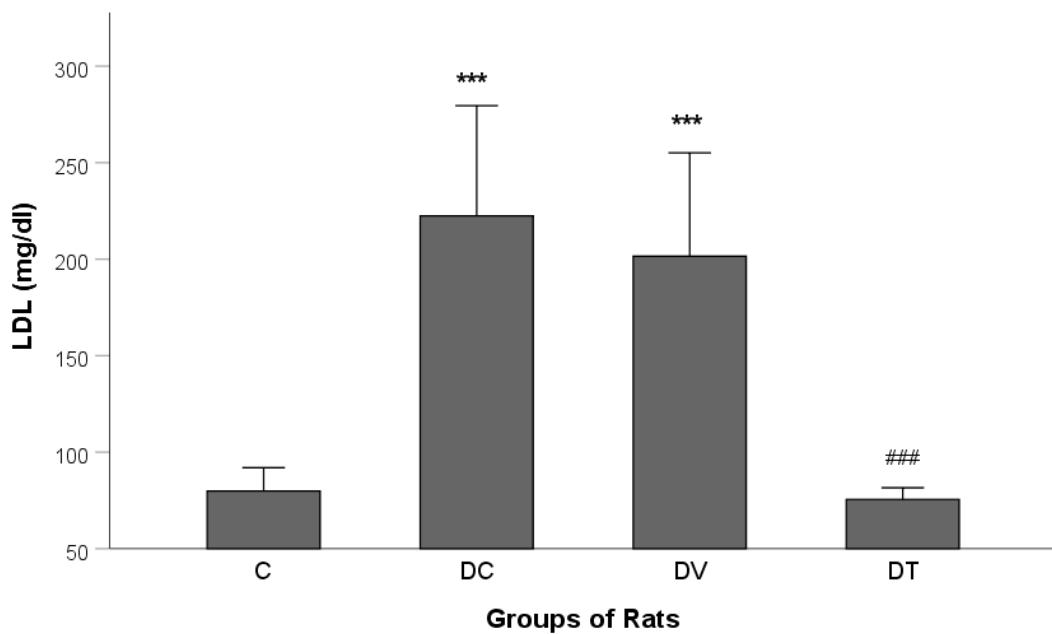


Fig 4. Mean LDL levels for the different groups of rats after 8 weeks of the study

Data were presented as mean \pm SD. LDL (low-density lipoproteins) mg/dl. C = Control rats (n=10), DC= Diabetic Control group (n=7), DV= Diabetic + Vehicle group (n=8), DT= Diabetic-DMH1 Treated group [5mg/kg q.o.d] (n=6). *** very highly significant ($P < 0.001$) against control group. ### very highly significant ($P < 0.001$) in comparison to Diabetic non-treated group (DC or DV)

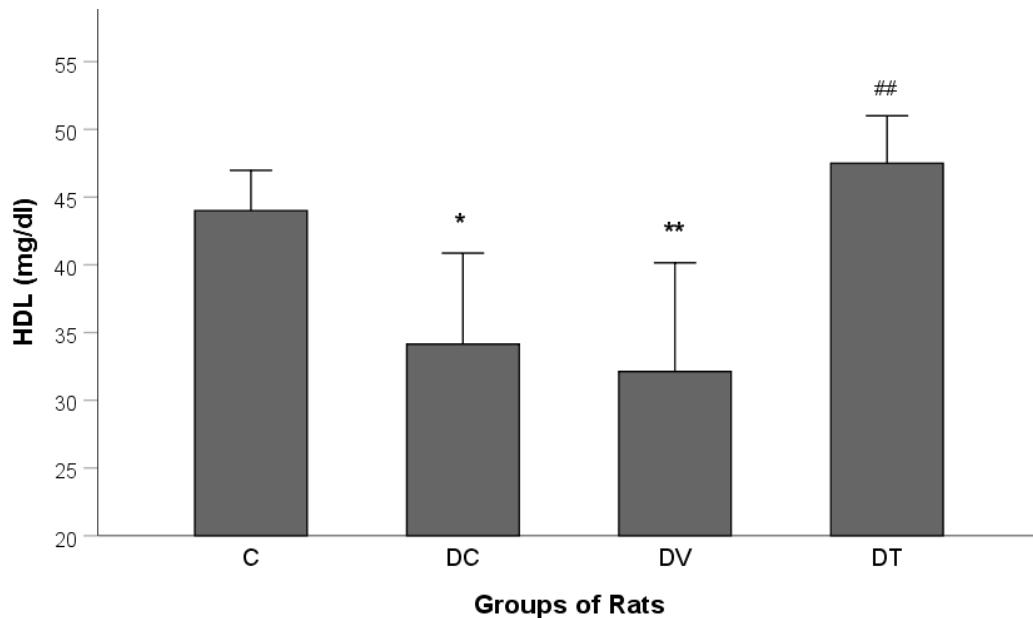


Fig. 5. Mean HDL levels for rats after 8 weeks of the study

Data were presented as mean \pm SD. HDL (high-density lipoproteins) mg/dl. C = Control rats (n=10), DC= Diabetic Control group (n=7), DV= Diabetic + Vehicle group (n=8), DT= Diabetic-DMH1 Treated group [5mg/kg q.o.d] (n=6). * significant ($P < 0.05$) against control group. ** highly significant ($P < 0.01$) against control group. ## highly significant ($P < 0.01$) compared to DC or DV group

4. DISCUSSION

Dyslipidemia is prevalent in diabetic patients and commonly linked to premature atherosclerotic cardiovascular outcomes. Additionally, there is a substantial evidence addressing the benefit of lipid-lowering drugs in reducing risk of CHD in patients with or without pre-existing cardiovascular condition [9]. However, more options to manage dyslipidemia and further clarification of potential mechanisms are critically needed. In the current study, effect of BMP inhibition using intraperitoneal administration of DMH1 was tested against diabetic-induced dyslipidemia in experimental rats.

In the current study, persistent hyperglycemia along the study period has been observed in all diabetic rats (DC, DV & DT). Therefore, this study stands with other studies to address the effectiveness of single dose STZ (60 mg/kg) as a diabetes-inducing agent in rats [29,30]. Remarkably, symptoms of polyuria and polydipsia were obvious and easily noticeable by the increase in daily consumption of chew and water. Thus, persistent untreated hyperglycemia might have led to severe dehydration which is the proposed cause of death for the 9 rats.

DMH1 is a small, potent and highly selective molecule. Originally, DMH1 was the designed analog of non-selective BMP receptor inhibitor (Dorsomorphin). However, unlike Dorsomorphin, DMH1 demonstrated higher stability, lower toxicity and the advantage of specifically inhibiting BMP-2 and BMP-4 induced Smad1/5/8 activation through Activin receptor-like kinase-1(ALK1), ALK2, and ALK3 receptors with negligible effect on ALK6 receptor and no effect on p38/MAPK signaling or vascular endothelial growth factor (VEGF) pathway [22]. Moreover, *in vivo* DMH1 administration was tested in few previous experiments which reported good tolerability and no sign of toxicity [24,27].

On the other hand, BMP family is a component of transforming growth factors beta (TGF- β) superfamily. It contained many members with pleiotropic effects on cellular development and fate, including adipogenesis. Notably, BMP-2 and BMP-4 were reported to activate the expression and phosphorylation of downstream signaling to initiate adipogenic commitment from a cell line of mesenchymal stem cells (MSCs), while knockdown of coregulator of BMP/Smad signaling (Smad4) resulted in disruption of this commitment [31]. Furthermore, a later study was

carried out to evaluate the association of BMP-4 levels with metabolic disorders. Interestingly, researchers found levels of BMP-4 were significantly correlated with waist circumference, body mass index, triglyceride (TG) and high-density lipoproteins (HDL) cholesterol [32].

In fact, dyslipidemia is known to associate with diabetes. In diabetic patients, the activation of triacylglycerol lipase enzyme led to more free fatty acids (FFA) mobilization from the adipose tissue with a consequent hepatic TG overproduction. Also, further increases in TG production and remnant cholesterol are due to suppressed activity of lipoprotein lipase (LPL) [33]. In the current study, dyslipidemia was found remarkable in both diabetic non-treated groups; total cholesterol, triglyceride, LDL showed significantly increased levels in DC & DV groups when compared to the control group. On the other hand, the DMH1-treated group showed readings within similar ranges to that of the normal rats. The current results are indicating a promising effect of DMH1 against dyslipidemia. Simultaneously, DMH1 administration resulted in a significant increase in HDL compared to the non-treated groups. So, both HDL and LDL cholesterol, as well as triglyceride and TC levels, were significantly and positively affected by DMH1.

Arguably, β -cyclodextrin (β CD), which (its derivative [HP β CD]) was used as a vehicle in the current study, was reported to decrease plasma lipid in the experimental rats [34]. β CD was claimed, due to its chemical structure (ring), to act as a sequestrant and slowly hydrolyzed in the large intestine and excreted intact in the feces when administered orally [34]. Contrarily, in the present study, HP β CD was given intraperitoneally (DV group) and did not demonstrate any lipid lowering effect. Other reports of sustained parenteral administration of β CD in animals were showing either increased [35] or unaffected plasma cholesterol [36]. Moreover, the administered dose in our study was much lower than the reported oral and parenteral doses. Furthermore, the cholesterol-lowering effect of an oral diet containing β CD was not uniformly in addition to a significant decrease in HDL cholesterol [34]. By contrast, the treated group in the current study showed a uniformly antihyperlipidemic effect with an increase in HDL cholesterol, an effect which is more plausible to be credited to DMH1.

In fact, our results are consistent with others who reported that inhibition of BMP would increase lipid efflux and reduce intracellular lipid accumulation. Almost ten years ago, LDN-193189, which is one of dorsomorphin structural analogs that has comparable DMH1 inhibitory property against BMP/Smad pathway, was investigated to explore its pharmacological effect in treating atherosclerosis. Results of the treated mice proved increasing lipid efflux and reduction in the formation of foam cells which have an important role in the occurrence and development of atherosclerosis [37]. Almost within the same year, another study has outlined BMP role in atherosclerosis using the same BMP inhibitor (LDN-193189) for 20 weeks with a high-fat diet fed to LDL receptor-deficient mice ($LDLR^{-/-}$), a commonly used model for atherosclerosis. Treatment with LDN-193189 has reduced cholesterol and LDL levels, but not HDL or triglyceride levels. Furthermore, the pharmacological BMP inhibition effectively decreased SMAD1/5/8 activation as well as inflammation, oxidative stress, atherosclerosis, vascular calcification and hepatic steatosis. Interestingly, the authors reported that early atherosclerotic lesions were distinguished by noticeable activation of the BMP signaling. Moreover, the cholesterol-lowering effect was appeared irrelevant to HMG CoA reductase or its hepatic gene expression, suggesting a novel therapeutic strategy for dyslipidemia [20].

In the current study, DMH1 lipid-lowering effects were significant and remarkable in total cholesterol, triglyceride and LDL levels. Furthermore, the average, as well as majority, of HDL cholesterol readings for the treated group showed a mild increase in comparison to the control group, however, this increase was not statistically significant. Although the current findings were based on the serum biomarkers solely, however, it is the first, as far as we know, to mark the antihyperlipidemic effect of DMH1. Also, the results can be useful to add to the existing evidence regarding the involvement of BMP in dyslipidemia and atherosclerosis as well as encouraging future investigations regarding DMH1 and its potential therapeutic applications.

5. CONCLUSION

In the current study, DMH1 revealed a significant antihyperlipidemic effect in animal model of diabetic dyslipidemia. This promising effect was favorable due to considerable decreases in TC, LDL and TG levels with, simultaneously,

remarkable increases in HDL levels. The current results can also reinforce the previous establishment of BMP/Smad involvement in dyslipidemia and atherosclerosis.

Furthermore, it can encourage future investigations for the underlying downstream signaling and the potential applicability or utilization of DMH1 in dyslipidemia-related experimental research.

DISCLAIMER

The products used for this research are commonly used products in many areas of research. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

CONSENT

It is not applicable.

ETHICAL APPROVAL

The current experiment was carried out based on the Good Clinical Practice (GCP) Guidelines and granted ethical approval from Research Ethics Committee (REC) at KAU (Reference No 546-19).

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Tseng CH. Mortality and Causes of Death in a National Sample of Diabetic Patients in Taiwan. *Diabetes Care*. 2004;27:1605.
2. Rao Kondapally Seshasai S, Kaptoge S, Thompson A, Di Angelantonio E, Gao P, Sarwar N, Whincup PH, Mukamal KJ, Gillum RF, Holme I, Njølstad I, Fletcher A, Nilsson P, Lewington S, Collins R, Gudnason V, Thompson SG, Sattar N, Selvin E, Hu FB, Danesh J, Emerging Risk Factors C. Diabetes mellitus, fasting glucose, and risk of cause-specific death. *The New England journal of medicine*. 2011;364:829-841.

3. Rawshani A, Sattar N, Franzén S, Rawshani A, Hattersley AT, Svensson A-M, Eliasson B, Gudbjörnsdóttir S. Excess mortality and cardiovascular disease in young adults with type 1 diabetes in relation to age at onset: a nationwide, register-based cohort study. *Lancet* (London, England). 2018;392:477-486.
4. Haffner SM, Lehto S, Rönnemaa T, Pyörälä K, Laakso M. Mortality from coronary heart disease in subjects with type 2 diabetes and in nondiabetic subjects with and without prior myocardial infarction. *N Engl J Med.* 1998;339: 229-234.
5. Evans JMM, Wang J, Morris AD. Comparison of cardiovascular risk between patients with type 2 diabetes and those who had had a myocardial infarction: cross sectional and cohort studies. *BMJ* (Clinical research ed.). 2002;324: 939-942.
6. Wannamethee SG, Shaper AG, Whincup PH, Lennon L, Sattar N. Impact of diabetes on cardiovascular disease risk and all-cause mortality in older men: influence of age at onset, diabetes duration, and established and novel risk factors. *Arch Intern Med.* 2011;171:404-410.
7. Goldberg IJ. Diabetic Dyslipidemia: Causes and Consequences. *The Journal of Clinical Endocrinology & Metabolism.* 2001;86:965-971.
8. Miller M, Stone NJ, Ballantyne C, Bittner V, Criqui MH, Ginsberg HN, Goldberg AC, Howard WJ, Jacobson MS, Kris-Etherton PM, Lennie TA, Levi M, Mazzone T, Pennathur S. Triglycerides and Cardiovascular Disease. *Circulation.* 2011; 123:2292-2333.
9. Schofield JD, Liu Y, Rao-Balakrishna P, Malik RA, Soran H. Diabetes Dyslipidemia. *Diabetes Therapy.* 2016;7:203-219.
10. Schwab KO, Doerfer J, Hecker W, Grulich-Henn J, Wiemann D, Kordonouri O, Beyer P, Holl RW. Spectrum and Prevalence of Atherogenic Risk Factors in 27,358 Children, Adolescents, and Young Adults With Type 1 Diabetes. *Diabetes Care.* 2006;29:218.
11. Ahmadizar F, Souverein P, de Boer A, Maitland-van der Zee AH. Undertreatment of hypertension and hypercholesterolaemia in children and adolescents with type 1 diabetes: long-term follow-up on time trends in the occurrence of cardiovascular disease, risk factors and medications use. *British Journal of Clinical Pharmacology.* 2018;84:776-785.
12. Kim G, DeSalvo D, Guffey D, Minard CG, Cephus C, Moodie D, Lyons S. Dyslipidemia in adolescents and young adults with type 1 and type 2 diabetes: a retrospective analysis. *International Journal of Pediatric Endocrinology.* 2020;2020:11.
13. Reaven G, Greenfield MS. Diabetic Hypertriglyceridemia: Evidence for Three Clinical Syndromes. *Diabetes.* 1981;30:66 - 75.
14. Hirano T. Pathophysiology of Diabetic Dyslipidemia. *Journal of atherosclerosis and thrombosis.* 2018;25:771-782.
15. Chen D, Zhao M, Mundy GR. Bone morphogenetic proteins. *Growth Factors.* 2004;22:233-241.
16. Dhore CR, Cleutjens JPM, Lutgens E, Cleutjens KBJM, Geusens PPM, Kitslaar PJEHM, Tordoir JHM, Spronk HMH, Vermeer C, Daemen MJAP. Differential Expression of Bone Matrix Regulatory Proteins in Human Atherosclerotic Plaques. *Arteriosclerosis, Thrombosis, and Vascular Biology.* 2001;21:1998-2003.
17. Ikeda K, Souma Y, Akakabe Y, Kitamura Y, Matsuo K, Shimoda Y, Ueyama T, Matoba S, Yamada H, Okigaki M, Matsubara H. Macrophages play a unique role in the plaque calcification by enhancing the osteogenic signals exerted by vascular smooth muscle cells. *Biochem Biophys Res Commun.* 2012;425:39-44.
18. Miller JD, Weiss RM, Heistad DD. Calcific aortic valve stenosis: methods, models, and mechanisms. *Circulation research.* 2011;108:1392-1412.
19. Nakagawa Y, Ikeda K, Akakabe Y, Koide M, Uraoka M, Yutaka K-t, Kurimoto-Nakano R, Takahashi T, Matoba S, Yamada H, Okigaki M, Matsubara H. Paracrine Osteogenic Signals via Bone Morphogenetic Protein-2 Accelerate the Atherosclerotic Intimal Calcification In Vivo. *Arteriosclerosis, Thrombosis, and Vascular Biology.* 2010;30:1908-1915.
20. Derwall M, Malhotra R, Lai CS, Beppu Y, Aikawa E, Seehra JS, Zapol WM, Bloch KD, Yu PB. Inhibition of Bone Morphogenetic Protein Signaling Reduces

- Vascular Calcification and Atherosclerosis. Arteriosclerosis, Thrombosis, and Vascular Biology. 2012;32:613-622.
21. Boström KI, Jumabay M, Matveyenko A, Nicholas SB, Yao Y. Activation of Vascular Bone Morphogenetic Protein Signaling in Diabetes Mellitus. Circulation Research. 2011;108:446-457.
 22. Hao J, Ho JN, Lewis JA, Karim KA, Daniels RN, Gentry PR, Hopkins CR, Lindsley CW, Hong CC. In Vivo Structure-Activity Relationship Study of Dorsomorphin Analogues Identifies Selective VEGF and BMP Inhibitors. ACS Chemical Biology. 2010;5:245-253.
 23. Ao A, Hao J, Hopkins CR, Hong CC. DMH1, a novel BMP small molecule inhibitor, increases cardiomyocyte progenitors and promotes cardiac differentiation in mouse embryonic stem cells. PLoS One. 2012;7:e41627.
 24. Hao J, Lee R, Chang A, Fan J, Labib C, Parsa C, Orlando R, Andresen B, Huang Y. DMH1, a Small Molecule Inhibitor of BMP Type I Receptors, Suppresses Growth and Invasion of Lung Cancer. PLOS ONE. 2014;9:e90748.
 25. Lin T, Wang X-L, Zettervall SL, Cai Y, Guzman RJ. Dorsomorphin homologue 1, a highly selective small-molecule bone morphogenetic protein inhibitor, suppresses medial artery calcification. Journal of vascular surgery. 2017;66: 586-593.
 26. Ren W, Zhao C, Wang Y, Fang Y, Huang Z, Chen W, Wang L, Hu W, Wang K, Ni L. Ramipril can alleviate the accumulation of renal mesangial matrix in rats with diabetic nephropathy by inhibiting insulin-like growth factor-1. Acta Cir Bras. 2019;34: e20190010000007.
 27. Ali JL, Lagasse BJ, Minuk AJ, Love AJ, Moraya AI, Lam L, Arthur G, Gibson SB, Morrison LC, Werbowetski-Ogilvie TE, Fu Y, Nachtigal MW. Differential cellular responses induced by dorsomorphin and LDN-193189 in chemotherapy-sensitive and chemotherapy-resistant human epithelial ovarian cancer cells. Int J Cancer. 2015; 136:E455-469.
 28. Loftsson T, Moya-Ortega MD, Alvarez-Lorenzo C, Concheiro A. Pharmacokinetics of cyclodextrins and drugs after oral and parenteral administration of drug/cyclodextrin complexes. J Pharm Pharmacol. 2016;68:544-555.
 29. Xu L, Shen P, Bi Y, Chen J, Xiao Z, Zhang X, Wang Z. Danshen injection ameliorates STZ-induced diabetic nephropathy in association with suppression of oxidative stress, pro-inflammatory factors and fibrosis. International Immunopharmacology. 2016;38:385-394.
 30. Du N, Xu Z, Gao M, Liu P, Sun B, Cao X. Combination of Ginsenoside Rg1 and Astragaloside IV reduces oxidative stress and inhibits TGF-β1/Smads signaling cascade on renal fibrosis in rats with diabetic nephropathy. Drug design, development and therapy. 2018;12: 3517-3524.
 31. Huang H, Song T-J, Li X, Hu L, He Q, Liu M, Lane MD, Tang Q-Q. BMP signaling pathway is required for commitment of C3H10T1/2 pluripotent stem cells to the adipocyte lineage. Proceedings of the National Academy of Sciences. 2009; 106:12670.
 32. Son J-W, Kim M-K, Park Y-M, Baek K-H, Yoo S-J, Song K-H, Son HS, Yoon K-H, Lee WC, Cha B-Y, Son H-Y, Kwon H-S. Association of serum bone morphogenetic protein 4 levels with obesity and metabolic syndrome in non-diabetic individuals. Endocrine Journal. 2011; 58:39-46.
 33. Kawanami D, Matoba K, Utsunomiya K. Dyslipidemia in diabetic nephropathy. Renal Replacement Therapy. 2016;2:16.
 34. Favier ML, Rémesy C, Moundras C, Demigné C. Effect of cyclodextrin on plasma lipids and cholesterol metabolism in the rat. Metabolism. 1995;44:200-206.
 35. Irie T, Fukunaga K, Garwood MK, Carpenter TO, Pitha J, Pitha J. Hydroxypropylcyclodextrins in parenteral use. II: Effects on transport and disposition of lipids in rabbit and humans. J Pharm Sci. 1992;81:524-528.
 36. Zimmer S, Grebe A, Bakke SS, Bode N, Halvorsen B, Ulas T, Skjelland M, De Nardo D, Labzin LI, Kerksiek A, Hempel C, Heneka MT, Hawthurst V, Fitzgerald ML, Trebicka J, Björkhem I, Gustafsson J-Å, Westerterp M, Tall AR, Wright SD, Espenvik T, Schultzze JL, Nickenig G, Lütjohann D, Latz E. Cyclodextrin promotes atherosclerosis regression via

- macrophage reprogramming. Science translational medicine. 2016;8: 333ra350-333ra350.
37. Saeed O, Otsuka F, Polavarapu R, Karmali V, Weiss D, Davis T, Rostad B, Pachura K, Adams L, Elliott J, Taylor WR, Narula J, Kolodgie F, Virmani R, Hong CC, Finn AV. Pharmacological suppression of hepcidin increases macrophage cholesterol efflux and reduces foam cell formation and atherosclerosis. Arterioscler Thromb Vasc Biol. 2012;32:299-307.

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